The Journal of Physiology

https://jp.msubmit.net

JP-RP-2021-282282R2

Title: The Influence of Maturation on Exercise-Induced Cardiac Remodelling and Haematological Adaptation

Authors: Dean Perkins
Jack Talbot
Rachel Lord
Tony Dawkins
Aaron Baggish
Abbas Zaidi
Orhan Uzun
Kelly Mackintosh
Melitta McNarry
Stephen-Mark Cooper
Rhodri Lloyd
Jon Oliver
Rob Shave
Mike Stembridge

Author Conflict: No competing interests declared

Author Contribution: Dean Perkins: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final

Disclaimer: This is a confidential document.

approval of the version to be published Jack Talbot: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Rachel Lord: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Tony Dawkins: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Aaron Baggish: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Abbas Zaidi: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Orhan Uzun: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Kelly Mackintosh: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Melitta McNarry: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Stephen-Mark Cooper: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Rhodri Lloyd: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Jon Oliver: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Rob Shave: Conception or design of the work; Acquisition or analysis or interpretation of data for the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published Mike Stembridge: Conception or design of the work; Acquisition or analysis or interpretation of data for

the work; Drafting the work or revising it critically for important intellectual content; Final approval of the version to be published; Agreement to be accountable for all aspects of the work

Running Title: Maturation and endurance training adaptation

Dual Publication: No

Funding: Dean's PhD research bursary, Cardiff Metropolitan University: Dean Perkins, n/a

Disclaimer: This is a confidential document.

1 The Influence of Maturation on Exercise-Induced Cardiac Remodelling and

2 Haematological Adaptation

- 3 Dean R Perkins^a, Jack S Talbot^a, Rachel N Lord^a, Tony G Dawkins^{a,h}, Aaron L
- 4 Baggish^b, Abbas Zaidi^c, Orhan Uzun^c, Kelly A Mackintosh^d, Melitta A McNarry^d,
- 5 Stephen-Mark Cooper^a, Rhodri S Lloyd^{e,f,g}, Jon L Oliver^{e,f}, Rob E Shave^h, and Mike
- 6 Stembridge^a
- ^a Cardiff School of Sport and Health Sciences, Cardiff Metropolitan University,
- 8 Cardiff, United Kingdom.
- ^b Cardiovascular Performance Program, Massachusetts General Hospital, Boston.
- ^c University Hospital of Wales, Cardiff, United Kingdom.
- ^d Applied Sports, Technology, Exercise and Medicine (A-STEM) Research Centre,
- 12 Swansea University, Swansea, United Kingdom.
- ^e Youth Physical Development Centre, Cardiff Metropolitan University, Cardiff, United
- 14 Kingdom
- ^f Sports Performance Research Institute New Zealand, AUT University, Auckland,
- 16 New Zealand
- 17 ⁹ Centre for Sport Science and Human Performance, Waikato Institute of
- 18 Technology, Waikato, New Zealand
- 19 h Centre for Heart, Lung and Vascular Health, School of Health and Exercise
- 20 Sciences, University of British Columbia Okanagan, Kelowna, Canada.

21 Corresponding Author

- 22 Mike Stembridge
- 23 Address for correspondence: Cardiff School of Sport and Health Sciences, Cardiff
- 24 Metropolitan University, Cyncoed Campus, Cyncoed Road, Cardiff, UK.

25 Email: <u>mstembridge@cardiffmet.ac.uk</u>

27

Keywords: echocardiography, puberty, paediatric, haematology, endurance training.

Key points

28

35

36

37

38

39

- It has long been hypothesised that cardiovascular adaptation to endurance training is augmented following puberty.
- We investigated whether differences in cardiac and haematological variables
 exist, and to what extent, between endurance-trained *vs.* untrained, pre- and
 post-peak height velocity (PHV) children, and how these central factors relate to
 maximal oxygen consumption.
 - Using echocardiography to quantify left ventricular (LV) morphology and carbon monoxide rebreathing to determine blood volume and haemoglobin mass, we identified that training-related differences in LV morphology are evident in pre-PHV children, with haematological differences also observed between pre-PHV girls. However, the breadth and magnitude of cardiovascular remodelling was more pronounced post-PHV.
- Cardiac and haematological measures provide significant predictive models for maximal oxygen consumption ($\dot{V}O_{2max}$) in children that are much stronger post-PHV, suggesting that other important determinants within the oxygen transport chain could account for the majority of variance in $\dot{V}O_{2max}$ before puberty.

Abstract

45

Cardiovascular and haematological adaptations to endurance training facilitate 46 greater maximal oxygen consumption (VO_{2max}), and such adaptations maybe 47 augmented following puberty. Therefore, we compared left ventricular (LV) 48 morphology (echocardiography), blood volume, haemoglobin (Hb) mass (CO-49 rebreathe) and $\dot{V}O_{2max}$ in endurance-trained and untrained boys (n=42, age=9.0-17.1 50 years, $\dot{V}O_{2max}$ =61.6±7.2 mL·kg·min, and n=31, age=8.0-17.7 years, $\dot{V}O_{2max}$ =46.5±6.1 51 mL·kg·min, respectively) and girls (n=45, age=8.2-17.0 years, $\dot{V}O_{2max}=51.4\pm5.7$ 52 mL·kg·min and n=36, age=8.0-17.6 years, $\dot{V}O_{2max}=39.8\pm5.7$ mL·kg·min, respectively). 53 Pubertal stage was estimated via maturity offset, with participants classified as pre-54 or post-peak height velocity (PHV). Pre-PHV, only a larger LV end-diastolic 55 volume/lean body mass (EDV/LBM) for trained boys (+0.28 mL·kg^{LBM}, P=0.007) and 56 a higher Hb mass/LBM for trained girls (+1.65 g·kg^{LBM}, P=0.007) were evident 57 compared to untrained controls. Post-PHV, LV mass/LBM (boys:+0.50 q·kqLBM, 58 P=0.0003; girls:+0.35 g·kg^{LBM}, P=0.003), EDV/LBM (boys:+0.35 mL·kg^{LBM}, 59 P<0.0001; girls:+0.31 mL·kgLBM, P=0.0004), blood volume/LBM (boys:+12.47 60 mL·kg^{LBM}, P=0.004; girls:+13.48 mL·kg^{LBM}, P=0.0002.) and Hb mass/LBM 61 (boys:+1.29 g·kg^{LBM}, P=0.015; girls:+1.47 g·kg^{LBM}, P=0.002) were all greater in 62 trained vs. untrained groups. Pre-PHV, EDV (R²_{adi}=0.224, P=0.001) in boys, and Hb 63 mass and interventricular septal thickness (R^2_{adi} =0.317, P=0.002) in girls partially 64 accounted for the variance in VO_{2max}. Post-PHV, stronger predictive models were 65 evident via the inclusion of LV wall thickness and EDV in boys ($R^2_{adi}=0.608$, 66 P<0.0001), and posterior wall thickness and Hb mass in girls ($R^2_{adi}=0.490$, 67 P<0.0001). In conclusion, cardiovascular adaptation to exercise training is more 68

pronounced post-PHV, with evidence for a greater role of central components for oxygen delivery.

Introduction

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

Cardiovascular adaptations to endurance training facilitate enhanced oxygen delivery while minimising cardiac work, in part setting the upper limit for endurance exercise performance (Lundby et al., 2017). These adaptations to the central components of the oxygen transport chain include cardiac remodelling, which enhances stroke volume (Morganroth et al., 1975; Pluim et al., 2000), and an expansion in haemoglobin (Hb) volume, increasing oxygen carrying capacity in the blood (Remes, 1979; Montero et al., 2017). In adults, cardiac and haematological adaptations to endurance training often occur concomitantly (Montero et al., 2015; Skattebo et al., 2020), although this is not always the case (Arbab-Zadeh et al., 2014). An enhanced circulating haematological volume with endurance training further stimulates cardiac remodelling via an increased ventricular filling pressure (Morganroth et al., 1975; Prior & La Gerche, 2012). Therefore, cardiac and haematological adaptations are not only key variables in determining maximal oxygen consumption, but cardiac remodelling may also be dependent on the extent and timing of haematological expansion in response to training. Nearly 40 years ago, it was hypothesised that cardiovascular training adaptations were absent in children before puberty due to low levels of sex- and growth-related hormones that increase substantially following puberty (Katch, 1983), particularly in boys (Wood et al., 2019). During adolescence, sex- and growth-related hormones result in a peak rate of lean tissue growth around the timing of peak height velocity (PHV) (Iuliano-Burns et al., 2001; Wood et al., 2019), and have also been associated with cardiovascular adaptation to exercise (Marsh et al., 1998; Neri Serneri et al.,

2001; Hero et al., 2005). Indeed, adult female athletes, who will naturally experience lower androgen levels, demonstrate less pronounced left ventricular (LV) hypertrophy in response to chronic endurance training in comparison to their male counterparts (Pelliccia et al., 1996). Despite lower growth-related hormone levels, a high $\dot{V}O_{2max}$ has been observed in pre-pubertal endurance-trained children (Mayers & Gutin, 1979; Nottin et al., 2002), and a recent meta-analysis demonstrated cardiac hypertrophy in athletes across the adolescent spectrum (McClean et al., 2018). However, LV hypertrophy was less prevalent in younger athletes, and evidence for training-related haematological adaptations in pre-pubertal children is sparse, with very few having investigated the area (Prommer et al., 2018). Therefore, speculation remains around whether puberty provides a window of opportunity for enhanced training-induced cardiovascular adaptation. If this is the case, haematological expansion with training around puberty could act as a physiological stimulus for enhanced cardiac remodelling compared with pre-puberty (Prior & La Gerche, 2012). VO_{2max} responses with training are similar between pre- and post-pubertal groups (Baquet et al., 2003; Runacres et al., 2019). Given that cardiovascular training adaptations may differ between these stages of maturation, the relative contributions of the central components of oxygen transport are likely to be different. It was therefore hypothesised that: (i) LV morphology and haematological components would be greater in all endurance-trained vs. untrained groups, but the magnitude of difference would be greater in post-, compared with pre-PHV cohorts; (ii) that blood volume would have a stronger relationship with LV end-diastolic volume post- vs. pre-PHV in boys and girls; and (iii) the variance in aerobic exercise capacity would be accounted for by both cardiac and haematological variables, with an increased contribution from these central components post-PHV. This study therefore aimed to:

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

(i) investigate whether there are any differences in cardiac and haematological variables by training status and, if so, whether the magnitude differs between pre- and post-PHV children; (ii) examine whether blood volume is associated with end-diastolic volume pre- and post-PHV; and (iii) identify the proportion of aerobic exercise capacity that can be accounted for by cardiac and haematological parameters pre- and post-PHV.

Methods

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

Ethical approval

The study was approved by the Cardiff Metropolitan University Natural Sciences Research Ethics Sub-committee (PGR-1339). Parents or guardians provided written informed consent and children provided written informed assent to participate in the study, which conformed to the ethical standards of the *Declaration of Helsinki*, except for registration in a database.

Study participants

A total of n = 163 participants were recruited. Participants were excluded due to failing to complete all measurements (n = 3), or failing to meet our cohort health or physical activity criteria (n = 6). Based on self- and parental-reported exercise training and physical activity, n = 154 participants were assigned to either endurance-trained (boys: n = 42, age = 9.0-17.1 years; girls: n = 45, age = 8.2-17.0 years) or untrained (boys: n = 31, age = 8.0-17.7 years; girls: n = 36, age = 8.0-17.6 years) groups. Criteria to be included within the endurance-trained group were to be undertaking at least three hours of structured endurance exercise-training per week for ≥12 months with an endurance sports club (cycling, swimming, long-distance running, or triathlon), and competing in their respective sport. This was in addition to meeting the UK minimum physical activity guidelines of at least 60 minutes of moderate intensity physical activity per day across the week (Department of Health and Social Care, 2019). Training histories and typical weekly volumes were reported by participants and confirmed by their parents. Untrained individuals were defined as not meeting the UK minimum physical activity guidelines (Department of Health and Social Care, 2019). All participants were reported to be healthy, normotensive, nonsmokers, free from any known cardiac or systemic diseases and were deemed not obese according to age- and sex-specific body mass index (BMI) cut-offs of the International Obesity Task Force criteria (Cole *et al.*, 2000).

Experimental design

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

Participants visited the laboratory on two occasions. Parents or guardians were asked to ensure their child refrained from heavy exercise and caffeine consumption 12 hours prior, and had not eaten a heavy meal within three hours prior to arrival.

During the first laboratory visit, body mass, height and sitting height were measured, with leg length then derived from height minus sitting height. These variables were then used to estimate maturity using sex-specific equations (Mirwald et al., 2002). As per the original recommendation of Mirwald et al. (2002), the maturity offset was used to categorise participants as pre-PHV or post-PHV, depending on whether the value was below or above zero, respectively. The equation has a typical error of 0.5 years; however, the accuracy of the prediction improves the closer participants are to PHV, making incorrect categorisation of our participants less likely. Moreover, the equation was found to be stable from -1 - +2 years predicted PHV (Koziel & Malina, 2018). Age from predicted PHV was used as a surrogate measure of puberty due to the non-invasive nature of the maturity offset measurement. Additionally, given that it relates to the point of maximal growth, it is the key stage of interest due to the associated growth-related hormones driving this process (Wood, 2019) and thus, potentially driving cardiac growth. Resting blood pressure was measured following 10 minutes supine rest using an automated sphygmomanometer (Omron Healthcare, Hoofddorp, Netherlands). $\dot{V}O_{2max}$ and maximal heart rate (HR_{max}) were assessed during a cardiopulmonary exercise test on an upright cycle ergometer (Lode, Excalibur, Groningen, Netherlands). Body composition, resting echocardiography and carbon monoxide (CO)-rebreathing measures were obtained during the second laboratory visit.

Experimental measures

Cardiorespiratory fitness

Participants completed an incremental ramp protocol on a cycle ergometer (Lode Excalibur; Groningen, The Netherlands) with ventilatory gas exchange measures for $\dot{V}O_2$, using a breath-by-breath gas analysis system (Jaeger, Oxycon Pro, Warwickshire, UK). Incremental workload increments were determined by stature and training status (Ellis *et al.*, 2017) and began subsequent to a three-minute warm up cycling at 10 watts. For trained and untrained participants >150 cm, the incremental workloads were 25 and 20 watts per minute, respectively; 125-149.9 cm, were 20 and 15 watts, respectively; and 110-124.9 cm, were 15 and 10 watts, respectively. Participants cycled at 75-85 rpm until they were unable to continue, despite strong verbal encouragement. This was followed by 15 minutes of seated rest before a constant-load supramaximal verification test at 105% of achieved peak power output to verify that $\dot{V}O_{2max}$ was achieved as described by Bhammar *et al.* (2017). $\dot{V}O_{2max}$ was accepted as the highest 30-second average value attained from either the ramp incremental test or the supramaximal verification test.

Body composition

Body fat percentage was derived from the measurement of skinfold thickness and validated, youth-specific equations, with a typical error of 3.6 and 3.9 for boys and girls, respectively (Slaughter *et al.*, 1988). Body fat mass and lean body mass (LBM) were calculated from the body fat percentage and total body mass.

Resting echocardiography

197

After 10 minutes supine rest, echocardiography was performed with a Vivid E9 198 system (GE Vingmed Ultrasound, Horten, Norway) using a 1.5 – 4 MHz transducer. 199 Two-dimensional images from the parasternal and apical acoustic windows were 200 attained with participants in the left lateral decubitus position. Images were stored 201 digitally for offline data analysis (Echopac, GE medical, Horton, Norway) by the 202 principal researcher (DRP). 203 LV mass was calculated using the area-length method (Lang et al., 2015). Relative 204 wall thickness was calculated as (posterior wall thickness (LVPWd) + interventricular 205 septal thickness (IVSd))/(LV internal diameter at end-diastole (LVIDd)). LV end-206 diastolic volume (EDV), and LV end-systolic volume (ESV) were calculated using the 207 208 biplane modified Simpson's technique. Stroke volume (SV) was calculated as EDV-ESV and cardiac output (Q) was then calculated as a product of SV and heart rate 209 (HR) taken from the ultrasound electrocardiograph. All measurements are presented 210 as absolute and scaled values where appropriate. Where scaling has been 211 implemented, linear measures were scaled to height and three-dimensional 212 measures were scaled to LBM in a dimensionally consistent manner (Dewey et al., 213 2008). This approach was chosen over an allometric approach due to the difficulty in 214 calculating a common scaling exponent from our relatively small sample size, and 215 the lack of published exponents across maturational groups. Intra-observer 216 coefficient of variation for LV morphology variables were EDV: 4.2%; ESV: 6.7%; SV: 217 4.5%; IVSd: 8.2%; LVPWd: 6.3%; and LVIDd: 3.5%. 218

Carbon monoxide rebreathing

Haematological data were determined using the optimised carbon monoxide (CO)rebreathe method as previously described (Schmidt & Prommer, 2005), after 15 minutes in a sitting position. Prior to commencing the procedure, participants were familiarised with the equipment (SpiCO, Blood tec GmbH, Bayreuth, Germany) and the rebreathing protocol. A nose clip was fitted to participants, and after exhaling, they positioned the spirometer with a 5-liter reservoir bag of pure oxygen attached ready for rebreathing. Participants were instructed to fully inhale, whilst a CO-bolus was administered, before holding a full lung volume for 10 seconds. Participants then continued rebreathing the CO and O2 balance through the spirometer until two minutes. Upon completion of rebreathing, participants fully exhaled into the bag before valve closure to enable quantification of unabsorbed CO using a portable CO analyser (Dräger Pac 3500; Dräger Safety, Lübeck, Germany). The calculated CObolus was reduced compared to the adult dose from 0.8-1.2 mL kg to 0.4-0.8 mL kg for our paediatric participants, as per previous recommendations (Prommer & Schmidt, 2007). Fingertip capillary blood samples were acquired before and after two minutes of CO-rebreathing to determine haematocrit (Hct), Hb concentration and the percentage of carboxyhaemoglobin (ABL80, Radiometer, Crawley, UK). Expired CO was also quantified prior to rebreathing and at four minutes following the onset of rebreathing using a portable CO analyser (Dräger Pac 3500; Dräger Safety, Lübeck, Germany). The reliability for the CO-rebreathe protocol with the current investigator was assessed from a paediatric subgroup of six boys and six girls. Two sets of haematological data were obtained separated by two to seven days. Our intraobserver coefficients of variation for Hb mass and blood volume in a paediatric population were 2.1% and 3.2%, respectively.

Statistical analysis

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

Data are expressed as means ± standard deviations (SD), unless stated otherwise. To analyse how well matched pre- and post-PHV groups of the same sex and training status were, independent samples t-tests were used to assess reported training volume and history between trained groups. To explore the differences between trained and untrained participants, at pre- and post-PHV, two-way ANOVAs with training and maturity status as the fixed factors were run independently for boys and girls. Independent samples t-tests were used to identify differences where there was a significant main effect. Effect sizes (Cohen's d) were calculated to assess the magnitude of any group differences. As per convention, effect sizes of 0.2, 0.5, 0.8 and 1.2 were accepted as small, medium, large and very large, respectively (Cohen, 1988; Sawilowsky, 2009). Relationships between blood volume and EDV, pre- and post-PHV for boys (n = 35 and 33 included, respectively) and girls (n = 33 and 39 included, respectively) were assessed using linear regression analysis with pooled trained and untrained data. Trained and untrained data were pooled for these analyses to explore the relationship between blood volume and EDV across a range of fitness levels to identify whether these relationships differ between pre- and post-PHV. To identify the proportion of relative VO_{2max} (mL·kg^{LBM}·min) contributed to by cardiac and haematological variables for each pre- and post-PHV group, trained and untrained pooled relative data were converted to z-scores and bivariate relationships were identified with Pearson's correlation coefficients. Variables with high multicollinearity (r > 0.85 and variance inflation factor (VIF) > 10) were removed from subsequent analyses. The remaining variables associated with VO_{2max} were entered into stepwise multiple linear regression analyses. Statistical analyses were performed using the Statistical Package for Social Science Software (version 24,

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

- Chicago, IL) and GraphPad (Prism Version 8.1.1, GraphPad Software, San Diego,
- 270 CA), with α set a priori as 0.05.

Results

272

290

291

292

293

294

295

296

273 Training and physical activity characteristics

Trained groups were recruited from either cycling, swimming, running or triathlon 274 clubs. The proportion of participants from each of these respective sports were as 275 follows (% from cycling/swimming/running/triathlon): pre-PHV trained bovs 276 (56/21/17/4%); post-PHV trained boys (68/0/5/26%); pre-PHV trained girls 277 (27/36/13/22%); and post-PHV girls (39/21/21/17%). Weekly endurance training 278 279 volume was not significantly different between pre- and post-PHV trained boys (8.6 ± 2.7 vs. 9.9 \pm 2.7 hrs·wk, P = 0.125), whereas it was lower in pre-compared with 280 post-PHV trained girls (6.0 \pm 2.5 vs. 8.9 \pm 3.6 hrs·wk, P = 0.003). Weekly strength 281 training volumes were low across all groups (Trained boys: pre-PHV, 0.1 ± 0.3 282 hrs·wk; post-PHV, 0.5 ± 0.7 hrs·wk; trained girls: pre-PHV, 0.4 ± 0.6 hrs·wk; post-283 PHV, 0.3 ± 0.6 hrs wk). As expected, years of training were lower in pre-compared 284 with post-PHV trained groups, irrespective of sex (3.8 \pm 1.5 vs. 6.0 \pm 2.8 years, P =285 0.002, and 2.6 \pm 1.5 vs. 4.4 \pm 2.3 years, P = 0.003, for boys and girls, respectively). 286 Untrained participants were undertaking a small amount weekly of physical activity 287 (Untrained boys: pre-PHV, 1.1 ± 0.9 hrs·wk; post-PHV, 0.9 ± 1.1 hrs·wk; untrained 288 girls: pre-PHV, 1.0 ± 0.9 hrs·wk; post-PHV, 0.6 ± 0.9 hrs·wk). 289

Participant characteristics and cardiorespiratory fitness

There were no differences in maturity offset, height, body mass or LBM between trained and untrained groups at either pre- or post-PHV (Table 1). Further, no differences were found for systolic or diastolic blood pressure between trained and untrained groups either pre- or post-PHV, for boys or girls. As expected, endurance-trained boys and girls had a higher cardiorespiratory fitness than their untrained counterparts both pre- and post-PHV.

Left ventricular dimensions and systolic function

LV dimensions are outlined for boys and girls in Tables 2 and 3, respectively and both LV mass and EDV relative to LBM are depicted in Figure 1. In pre-PHV children, no significant differences were found in wall thicknesses between trained and untrained groups, aside from a greater IVSd/height in trained girls. Post-PHV, both IVSd/height and LVPWd/height were greater in both trained groups vs. untrained. Relative wall thickness was greater in trained vs. untrained girls post-PHV, but no difference was observed pre-PHV, or between boys by training status irrespective of maturity. Pre-PHV, there was no significant difference in LV mass scaled to LBM between trained vs. untrained groups; however, a difference was found post-PHV with large and very large effect sizes for both boys and girls, respectively.

Irrespective of maturity status, EDV and SV normalised to LBM were higher in trained vs. untrained boys, with a greater effect size post-PHV. In contrast, there was no significant difference in EDV or SV normalised to LBM between trained vs. untrained girls pre-PHV, however, both were higher in trained girls post-PHV, compared with untrained.

Haematological parameters

Haematological variables are detailed for boys and girls in Tables 2 and 3, respectively, and both blood volume and Hb mass relative to LBM are depicted in Figure 1. There were no training-related differences in haematological variables between pre-PHV boys. In contrast, pre-PHV trained girls had a higher relative Hb mass, blood volume and plasma volumes than untrained girls. Post-PHV, trained boys and trained girls had higher relative Hb mass, blood volume and plasma

volume when compared with untrained controls. Post-PHV, effect sizes were larger between trained and untrained boys compared with pre-PHV for relative measures of Hb mass, blood volume and plasma volume. Effect sizes were larger between trained and untrained girls post-PHV for relative blood volumes compared with pre-PHV, but similar between pre- and post-PHV groups for other relative haematological variables.

Relationship between end-diastolic volume and blood volume

No relationship was observed in pre-PHV boys between EDV, and blood volume normalised for LBM ($R^2 = 0.051$, P = 0.193), but a small, significant relationship was found with post-PHV boys ($R^2 = 0.184$, P = 0.013) (Figure 2). Similarly, a weak relationship was found between EDV and blood volume normalised for LBM with pre-PHV girls ($R^2 = 0.124$, P = 0.045), with a stronger relationship found with post-PHV girls ($R^2 = 0.316$, P = 0.0002).

Independent relationships with \dot{V} O_{2max}

Bivariate associations with $\dot{V}O_{2max}$ for cardiac structural and haematological variables are presented in Table 4. The only significant correlations identified for pre-PHV boys were ESV, EDV and SV (r=0.42-0.49, P=0.001-0.006). In post-PHV boys, significant correlations were found for IVSd, LVPWd, LVIDd, LV mass, ESV, EDV, SV, Hb mass and blood volume (r=0.41-0.69, P<0.0001-0.018). For pre-PHV girls, there were significant correlations between $\dot{V}O_{2max}$ and IVSd, LV mass, EDV, SV, Hb mass and blood volumes (r=0.35-0.49, P=0.004-0.034). In post-PHV girls, significant correlations were found for IVSd, LVPWd, LV mass, ESV, EDV, SV, Hb mass and blood volume (r=0.23-0.59, P<0.0001-0.023).

Multiple regression analysis

Multicollinearity of *z*-scores for relative variables were identified between LV volume measures for all groups, and haematological measures for pre- and post-PHV girls, and post-PHV boys. Multicollinear variables were removed as necessary prior to analyses. The only variable to contribute to a significant proportion of the variance in $\dot{V}O_{2max}$ for pre-PHV boys was EDV, which accounted for 22% of the variance. The variance in $\dot{V}O_{2max}$ was also accounted for by EDV, alongside IVSd and Hb mass for post-PHV boys, which significantly contributed 61% of the variance (Table 5). For pre-PHV girls, Hb mass and IVSd significantly contributed 32% of the variance in $\dot{V}O_{2max}$. Hb mass and LVPWd contributed a significant proportion of the variance in $\dot{V}O_{2max}$ for post-PHV girls, accounting for 49% of the variance. These models which account for the variance in $\dot{V}O_{2max}$ using *z*-scores are stronger post-PHV as demonstrated by greater adjusted R^2 values and smaller standard errors compared with pre-PHV groups, for boys and girls.

Discussion

In relation to our three hypotheses, the novel findings were: (i) cardiac and haematological differences between trained vs. untrained children appear more pronounced in post-PHV children compared to their pre-PHV counterparts, characterised by a larger magnitude of LV hypertrophy and higher blood volume in the older group; (ii) the relationship between blood volume and ventricular volumes was stronger post-PHV; and (iii) cardiac and haematological adaptations provide a substantially greater contribution to relative $\dot{V}O_{2max}$ post-PHV, suggesting a maturation-dependent shift towards the central components of oxygen delivery in the context of maximal oxygen consumption.

The influence of maturity on LV morphology with endurance training

It has long been speculated that puberty provides a window whereby cardiac adaptations to endurance exercise are enhanced due to the hormonal milieu at this stage of development (Katch, 1983; McClean *et al.*, 2018). In trained pre-PHV children, a larger LV volume in boys and greater interventricular wall thickness in girls was found, but no other evidence of remodelling. In contrast, a similar phenotype to the adult athlete's heart with greater LV mass, ventricular volumes and consistently thicker ventricular walls compared to untrained counterparts was found for the post-PHV group (Pluim *et al.*, 2000; Prior & La Gerche, 2012). Given the high training volume and $\dot{V}O_{2max}$ in our trained pre-PHV groups, this potentially suggests a limited capacity for exercise-induced cardiac remodelling compared to the adult heart. Previous research examining exercise-induced cardiac remodelling prior to the onset of puberty has found similar results to our study, with either LV dilation (Obert *et al.*, 1998; Obert *et al.*, 2001; Obert *et al.*, 2003) or increased wall thickness

(Geenen *et al.*, 1982; Ayabakan *et al.*, 2006; Larsen *et al.*, 2018) in isolation, rather than in combination. These isolated adaptations may reflect the beginning of phasic cardiac remodelling, similar to the adaptation process observed in adults (Weiner *et al.*, 2015).

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

In adult training studies, enhanced wall thickness or LV dilation have also been observed in isolation prior to an eventual LV eccentric hypertrophy (Arbab-Zadeh et al., 2014; Weiner et al., 2015). Arbab-Zadeh et al. (2014) found an initial increase in LV wall thickness during the first six to nine months of training in exercise naïve adults, with LV dilation observed thereafter. This is congruent with the present study, in which girls had an enhanced wall thickness pre-PHV and an increased volume post-PHV. Conversely, Weiner et al. (2015) observed LV dilation prior to increased wall thickness with training intensification in athletes, which is in accord with our observations between pre- and post-PHV cardiac adaptations in boys. The differential response in boys and girls could be explained by differences in training volume and intensity. For example, Arbab-Zadeh et al. (2014) demonstrated that lower training volumes and intensities lead to increased wall thicknesses, whereas high intensity and volume exercise results in volumetric adaptation. Pre-pubertal training studies have also demonstrated this with isolated wall thickness adaptation when a lesser training load was implemented (Larsen et al., 2018), compared with LV dilation alone when sessions are longer and completed at >80% maximal heart rate (Obert et al., 2003). In the current study, trained pre-PHV girls had a slightly, but significantly lower training volume than boys, which may explain the isolated wall thickness and LV dilation adaptations in each group, respectively. However, given that neither pre-PHV boys or girls presented with combined wall thickness and

volume adaptations, despite their extensive training volume, suggests that cardiac remodelling is likely limited prior to puberty.

The maturity related differences in LV mass could also be related to blood pressure, which increases from childhood to adolescence (Rosner *et al.*, 1993), as shown in the present data. Importantly though, resting blood pressures were similar between trained and untrained groups, regardless of maturity group. Although not measured in the current study, a more likely influence on differences in LV morphology is the systolic blood pressure response during exercise, which has a much stronger association with LV mass (Lauer *et al.*, 1992) and is greater in post-pubertal children (Wanne & Haapoja, 1988). This could indicate that although our post-PHV groups are undertaking a similar training volume, they likely experience a far greater afterload stimulus for remodelling.

The influence of maturation on haematological adaptations to endurance training

There was a difference in relative Hb mass and blood volume between trained and untrained boys post-PHV, but not pre-PHV. Haematological studies examining adaptations to endurance training in children and adolescents are sparse. However, similar to the present data in boys, Prommer *et al.* (2018) found that trained children under 12 years of age have no difference in these haematological components when compared with untrained counterparts. Continued monitoring of the trained group for a further 3.5 years revealed an exponential increase in Hb mass for boys after 12 years of age. Indeed, Prommer *et al.* (2018) found a relationship between Hb mass and LBM, but observed a 7% increase in Hb mass that was unrelated to body size and attributed to the effects of training. Although maturity status was not quantified, Prommer *et al.* (2018) speculated that the increase in Hb mass was directly related

to increased testosterone. Erythropoiesis has been shown to be upregulated during puberty (Krabbe et~al., 1978) and related directly to androgens (Hero et~al., 2005; Coviello et~al., 2008). This could explain the relative difference in haematological components between trained and untrained boys that exists post-PHV, but not pre-PHV in the current study. In contrast, the scaled differences in haematological components between trained and untrained girls are similar pre- and post-PHV, rather than widening post-PHV. It could be postulated that such findings are a result of the markedly lower increase in testosterone in girls compared with boys at puberty (Handelsman et~al., 2018). Indeed, Prommer et~al. (2018) also found that whereas boys had an exponential increase in Hb mass around 12 years of age, the trajectory for trained girls remained unchanged across the study period, but only a very small number of girls (n=4) were studied making definitive conclusions problematic.

Enhanced blood volume as a stimulus for post-PHV LV adaptation

It is well established that endurance training leads to cardiac remodelling in adults (Fagard, 2003). This adaptation is partly attributed to the training-related increases in blood volume (Green *et al.*, 1991) and the associated increase in preload (Colan, 1997). In the present study, a stronger relationship between ventricular volumes and blood volumes was evident post-PHV when circulating blood volume was significantly larger in trained vs. untrained adolescents. These data indicate that the increase in circulating volume could provide an enhanced volume challenge further driving LV remodelling with endurance training post-puberty.

Cardiac and haematological determinants of \dot{V} O_{2max} pre- and post-PHV

Cardiac and haematological attributes are known to underpin $\dot{V}O_{2max}$ in adults (La Gerche *et al.*, 2012; Montero *et al.*, 2015; Diaz-Canestro *et al.*, 2021), but there is a

paucity of data defining cardiovascular determinants of $\dot{V}O_{2max}$ in adolescents. This study found that pre-PHV, the only variables to significantly contribute towards the variance in $\dot{V}O_{2max}$ were EDV for boys, and Hb mass and IVSd for girls, highlighting that contributions to endurance performance in pre-pubertal children are potentially sex dependent. The isolated cardiac variable and absence of a haematological influence in pre-PHV boys could reflect the lack of testosterone before puberty (Wood et al., 2019), given its stimulatory effect on erythropoiesis (Hero et al., 2005) and its association with cardiac hypertrophy (Marsh et al., 1998). Our findings post-PHV support this, with Hb mass and IVSd also emerging as significant contributors alongside EDV to partially account for $\dot{V}O_{2max}$ in the more mature boys. Interestingly, and in contrast to this finding, Hb mass was identified to significantly contribute to some of the variance in $\dot{V}O_{2max}$ in pre-PHV girls, alongside IVSd, partially accounting the variance. Although paediatric data are sparse, adult haematological adaptation to training appears to be similar between males and females (Montero et al., 2017). However, females are known to have a blunted cardiac adaptation to endurance training compared with males (Howden et al., 2015), which may explain the reduced proportion of $\dot{V}O_{2max}$ that IVSd accounts for pre-PHV girls compared with Hb mass. Therefore, oxygen carrying capacity rather than maximal cardiac output may be of greater importance in accounting for the variance in $\dot{V}O_{2max}$ for pre-pubertal girls. Further research is required to understand the temporal nature of haematological and cardiac adaptations to long-term endurance training in pre-pubertal boys and girls.

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

We found the strength of the $\dot{V}O_{2max}$ predictive models to be weaker in pre-PHV groups compared to post-PHV groups for both boys and girls, despite comparable cardiorespiratory fitness. Therefore, central factors appear to be of less importance

in contributing towards the variance in aerobic exercise capacity pre-, compared with post-puberty. It is well documented that aerobic energy metabolism is the predominant energy pathway in pre-pubertal children (Ratel & Blazevich, 2017) with anaerobic contributions increasing with maturity (Van Praagh & Dore, 2002). Compared with adults, pre-pubertal children have enhanced muscle oxidative potential which has been attributed to a higher oxidative enzyme activity (Haralambie, 1982), increased mitochondrial density (Bell *et al.*, 1980) and improved clearance rates of H $^+$ ions (Ratel *et al.*, 2008). Given that central parameters impart a relatively small contribution to $\dot{V}O_{2max}$ in our pre-PHV groups, we speculate that these other important determinants within the oxygen transport chain could account for the majority of variance in pre-pubertal aerobic exercise capacity. However, we acknowledge that adding more variables to the models would likely alter the proportions of the variance in $\dot{V}O_{2max}$ that the significant contributors in the current study account for.

Limitations

Due to the cross-sectional design, we were unable to establish causality for training related adaptations, however cardiac adaptations to training pre-puberty (Obert *et al.*, 2003) and during adolescence (Churchill *et al.*, 2020) have been observed. We were also unable to control for the greater training histories in post-PHV groups, nor the slightly higher training volume in post-PHV trained girls, and thus we cannot discount the potential influence of these factors. However, after removing trained participants with the highest and lowest historical training volumes to match pre- and post-PHV trained groups on these variables, we ran subgroup analyses for our key outcome measures. Using these subgroups of our trained participants with n = 13 in each pre- and post-PHV group, compared with the same untrained groups, there

were no significant changes to our results. To completely account for these training histories and volumes, longitudinal training interventions, and ideally twin training interventions are required with a focus on the influence of maturation. Additionally, the absence of atrial and right ventricular data is acknowledged as a limitation and future research is required to characterise these variables with training pre- and post-puberty. We also recognise that the gold standard technique for cardiac structure is magnetic resonance imaging (Grothues et al., 2002). However, echocardiography is frequently used in the assessment of cardiac remodelling (Lang et al., 2015) and has been validated in children (Lopez et al., 2010). The aim of the current study was to identify how cardiac and haematology influence $\dot{V}O_{2max}$, but we acknowledge that additional measures would also contribute to the variance in VO_{2max}. Future studies should consider other central and peripheral determinants within the oxygen transport chain especially in pre-pubertal children. Finally, we acknowledge our indirect method of quantifying maturation and recognise that the assessment of skeletal maturity would have provided the most accurate measure (Lloyd et al., 2014). Additionally, direct measures of hormones would have enabled direct associations with our key outcome variables. However, given the circadian fluctuations of sex- and growth-related hormones, multiple measures during the day and night would have been required for an accurate representation (Gupta et al., 2000; Matchock et al., 2007). Therefore, we did not include these in order to avoid too many disruptive and invasive measures in our young paediatric cohort.

Translational perspective

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

522

523

524

525

526

527

528

Given that competitive youth athletes undertake high training volumes throughout their developmental years, it is important to identify how such loads may present upon clinical examination at different stages of maturity. Our findings suggest that when attempting to differentiate between physiological and pathological cardiac remodelling, stage of maturity should be considered alongside endurance training history. Critically, our data suggest that marked LV dilation and wall thickening is very uncommon pre-puberty and should be considered pathologic until proven otherwise. Continued endurance training throughout puberty would then be expected to lead to more pronounced LV wall thickening and dilation as a feature of normal adaptation in the young athlete's heart.

Conclusion

Some degree of cardiac remodelling and haematological adaptation to endurance training is evident before puberty but is more pronounced following puberty. As children progress from childhood through adolescence, we speculate there may be a shift in the balance from peripheral to central components to account for the majority of the variance in maximal of oxygen consumption. However, pre-pubertal children remain eminently trainable and capable of achieving high levels of aerobic fitness – albeit potentially through different mechanisms than their older counterparts.

Additional information

Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Competing interests

The authors have no competing interests to declare.

Author contributions

DRP, RSL, RES, JLO and MS contributed to the conception and design of the study. All authors were involved with the acquisition, analysis, or interpretation of data. DRP and MS drafted the manuscript, and all authors were involved in revising it critically for important intellectual content. All authors approved the final version of the manuscript.

Funding

None

Acknowledgements

We appreciate and thank all participants and their parents/guardians for being a part of this study. Thanks also goes to Kerry Owen and Jo Phillips for their assistance during the recruitment process. The work was conducted at Cardiff Metropolitan University and the authors are grateful for the laboratory assistance provided by the Physiology Technician team. We also thank Bryony Curry, Thomas Griffiths, Katie Noteman, Zavia Penn and Cory Richards for their contributions towards data collection for this study.

568 569 570	Arbab-Zadeh A, Perhonen M, Howden E, Peshock RM, Zhang R, Adams-Huet B, Haykowsky MJ & Levine BD. (2014). Cardiac remodeling in response to 1 year of intensive endurance training. Circulation 130, 2152-2161.
571 572 573	Ayabakan C, Akalin F, Mengutay S, Cotuk B, Odabas I & Ozuak A. (2006). Athlete's heart in prepubertal male swimmers. <i>Cardiol Young</i> 16, 61-66.
574 575 576	Baquet G, van Praagh E & Berthoin S. (2003). Endurance training and aerobic fitness in young people. Sports Med 33 , 1127-1143.
577 578 579	Bell RD, MacDougall JD, Billeter R & Howald H. (1980). Muscle fiber types and morphometric analysis of skeletal msucle in six-year-old children. <i>Med Sci Sports Exerc</i> 12 , 28-31.
580 581 582	Bhammar DM, Stickford JL, Bernhardt V & Babb TG. (2017). Verification of Maximal Oxygen Uptake in Obese and Nonobese Children. <i>Med Sci Sports Exerc</i> 49 , 702-710.
583 584 585	Churchill TW, Groezinger E, Loomer G & Baggish AL. (2020). Exercise-induced cardiac remodeling during adolescence. <i>Eur J Prev Cardiol</i> 27 , 2148-2150.
586 587 588	Cohen J. (1988). Statistical power analysis for the behavioral sciences. L. Erlbaum Associates, Hillsdale, N.J.
589 590 591	Colan SD. (1997). Mechanics of left ventricular systolic and diastolic function in physiologic hypertrophy of the athlete's heart. <i>Cardiol Clin</i> 15, 355-372.
592 593 594	Cole TJ, Bellizzi MC, Flegal KM & Dietz WH. (2000). Establishing a standard definition for child overweight and obesity worldwide: international survey. <i>BMJ</i> 320 , 1240-1243.
595 596 597 598	Coviello AD, Kaplan B, Lakshman KM, Chen T, Singh AB & Bhasin S. (2008). Effects of graded doses of testosterone on erythropoiesis in healthy young and older men. <i>J Clin Endocrinol Metab</i> 93 , 914-919.
599 600	Department of Health and Social Care. (2019). UK Chief Medical Officers' Physical Activity Guidelines.
601 602 603	Dewey FE, Rosenthal D, Murphy DJ, Jr., Froelicher VF & Ashley EA. (2008). Does size matter? Clinical applications of scaling cardiac size and function for body size. <i>Circulation</i> 117 , 2279-2287.
604 605 606	Diaz-Canestro C, Pentz B, Sehgal A & Montero D. (2021). Sex Differences In Cardiorespiratory Fitness Are Explained By Blood Volume And Oxygen Carrying Capacity. <i>Cardiovasc Res</i> .
607	

References

608 609 610	Ellis LA, Ainslie PN, Armstrong VA, Morris LE, Simair RG, Sletten NR, Tallon CM & McManus AM. (2017). Anterior cerebral blood velocity and end-tidal CO2 responses to exercise differ in children and adults. <i>Am J Physiol Heart Circ Physiol</i> 312 , H1195-H1202.
611 612	Fagard R. (2003). Athlete's heart. <i>Heart</i> 89, 1455-1461.
613 614 615 616	Geenen DL, Gilliam TB, Crowley D, Moorehead-Steffens C & Rosenthal A. (1982). Echocardiographic measures in 6 to 7 year old children after an 8 month exercise program. <i>Am J Cardiol</i> 49 , 1990-1995.
617 618 619	Green HJ, Sutton JR, Coates G, Ali M & Jones S. (1991). Response of red cell and plasma volume to prolonged training in humans. <i>J Appl Physiol (1985)</i> 70 , 1810-1815.
620 621 622 623 624	Grothues F, Smith GC, Moon JC, Bellenger NG, Collins P, Klein HU & Pennell DJ. (2002). Comparison of interstudy reproducibility of cardiovascular magnetic resonance with two-dimensional echocardiography in normal subjects and in patients with heart failure or left ventricular hypertrophy. <i>Am J Cardiol</i> 90 , 29-34.
625 626 627	Gupta SK, Lindemulder EA & Sathyan G. (2000). Modeling of circadian testosterone in healthy men and hypogonadal men. <i>J Clin Pharmacol</i> 40, 731-738.
628 629 630	Handelsman DJ, Hirschberg AL & Bermon S. (2018). Circulating Testosterone as the Hormonal Basis of Sex Differences in Athletic Performance. <i>Endocr Rev</i> 39 , 803-829.
631 632 633	Haralambie G. (1982). Enzyme activities in skeletal muscle of 13-15 years old adolescents. <i>Bull Eur Physiopathol Respir</i> 18, 65-74.
634 635 636	Hero M, Wickman S, Hanhijarvi R, Siimes MA & Dunkel L. (2005). Pubertal upregulation of erythropoiesis in boys is determined primarily by androgen. <i>J Pediatr</i> 146 , 245-252.
637 638 639 640	Howden EJ, Perhonen M, Peshock RM, Zhang R, Arbab-Zadeh A, Adams-Huet B & Levine BD. (2015). Females have a blunted cardiovascular response to one year of intensive supervised endurance training. <i>J Appl Physiol (1985)</i> 119 , 37-46.
641 642 643 644	Iuliano-Burns S, Mirwald RL & Bailey DA. (2001). Timing and magnitude of peak height velocity and peak tissue velocities for early, average, and late maturing boys and girls. <i>Am J Hum Biol</i> 13 , 1-8.
645 646	Katch VL. (1983). Physical conditioning of children. <i>J Adolesc Health Care</i> 3, 241-246.
647 648 649	Koziel SM & Malina RM. (2018). Modified Maturity Offset Prediction Equations: Validation in Independent Longitudinal Samples of Boys and Girls. <i>Sports Med</i> 48, 221-236.
650	

651 652 653	Krabbe S, Christensen T, Worm J, Christiansen C & Transbol I. (1978). Relationship between haemoglobin and serum testosterone in normal children and adolescents and in boys with delayed puberty. <i>Acta Paediatr Scand</i> 67 , 655-658.
654 655 656 657	La Gerche A, Burns AT, Taylor AJ, Macisaac AI, Heidbuchel H & Prior DL. (2012). Maximal oxygen consumption is best predicted by measures of cardiac size rather than function in healthy adults. <i>Eur J Appl Physiol</i> 112 , 2139-2147.
658 659 660 661 662 663 664	Lang RM, Badano LP, Mor-Avi V, Afilalo J, Armstrong A, Ernande L, Flachskampf FA, Foster E, Goldstein SA, Kuznetsova T, Lancellotti P, Muraru D, Picard MH, Rietzschel ER, Rudski L, Spencer KT, Tsang W & Voigt JU. (2015). Recommendations for cardiac chamber quantification by echocardiography in adults: an update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. <i>J Am Soc Echocardiogr</i> 28, 1-39 e14.
665 666 667 668	Larsen MN, Nielsen CM, Madsen M, Manniche V, Hansen L, Bangsbo J, Krustrup P & Hansen PR. (2018). Cardiovascular adaptations after 10 months of intense school-based physical training for 8- to 10-year-old children. <i>Scand J Med Sci Sports</i> 28 Suppl 1, 33-41.
669 670 671 672	Lauer MS, Levy D, Anderson KM & Plehn JF. (1992). Is there a relationship between exercise systolic blood pressure response and left ventricular mass? The Framingham Heart Study. <i>Ann Intern Med</i> 116 , 203-210.
673 674 675 676	Lloyd RS, Oliver JL, Faigenbaum AD, Myer GD & De Ste Croix MB. (2014). Chronological age vs. biological maturation: implications for exercise programming in youth. <i>J Strength Cond Res</i> 28 , 1454-1464.
677 678 679 680 681 682	Lopez L, Colan SD, Frommelt PC, Ensing GJ, Kendall K, Younoszai AK, Lai WW & Geva T. (2010). Recommendations for quantification methods during the performance of a pediatric echocardiogram: a report from the Pediatric Measurements Writing Group of the American Society of Echocardiography Pediatric and Congenital Heart Disease Council. <i>J Am Soc Echocardiogr</i> 23 , 465-495; quiz 576-467.
683 684 685	Lundby C, Montero D & Joyner M. (2017). Biology of VO2 max: looking under the physiology lamp. Acta Physiol (Oxf) 220, 218-228.
686 687 688	Marsh JD, Lehmann MH, Ritchie RH, Gwathmey JK, Green GE & Schiebinger RJ. (1998). Androgen receptors mediate hypertrophy in cardiac myocytes. <i>Circulation</i> 98, 256-261.
689 690 691	Matchock RL, Dorn LD & Susman EJ. (2007). Diurnal and seasonal cortisol, testosterone, and DHEA rhythms in boys and girls during puberty. <i>Chronobiol Int</i> 24 , 969-990.
692 693 694	Mayers N & Gutin B. (1979). Physiological characteristics of elite prepubertal cross-country runners. <i>Med Sci Sports</i> 11, 172-176.

696 697 698	McClean G, Riding NR, Ardern CL, Farooq A, Pieles GE, Watt V, Adamuz C, George KP, Oxborough D & Wilson MG. (2018). Electrical and structural adaptations of the paediatric athlete's heart: a systematic review with meta-analysis. <i>Br J Sports Med</i> 52 , 230.
699 700 701	Mirwald RL, Baxter-Jones AD, Bailey DA & Beunen GP. (2002). An assessment of maturity from anthropometric measurements. <i>Med Sci Sports Exerc</i> 34, 689-694.
702 703 704 705	Montero D, Breenfeldt-Andersen A, Oberholzer L, Haider T, Goetze JP, Meinild-Lundby AK & Lundby C. (2017). Erythropoiesis with endurance training: dynamics and mechanisms. <i>Am J Physiol Regul Integr Comp Physiol</i> 312 , R894-R902.
706 707 708 709 710	Montero D, Cathomen A, Jacobs RA, Fluck D, de Leur J, Keiser S, Bonne T, Kirk N, Lundby AK & Lundby C. (2015). Haematological rather than skeletal muscle adaptations contribute to the increase in peak oxygen uptake induced by moderate endurance training. <i>J Physiol</i> 593 , 4677-4688.
711 712 713	Morganroth J, Maron BJ, Henry WL & Epstein SE. (1975). Comparative left ventricular dimensions in trained athletes. <i>Ann Intern Med</i> 82, 521-524.
714 715 716 717	Neri Serneri GG, Boddi M, Modesti PA, Cecioni I, Coppo M, Padeletti L, Michelucci A, Colella A & Galanti G. (2001). Increased cardiac sympathetic activity and insulin-like growth factor-l formation are associated with physiological hypertrophy in athletes. <i>Circ Res</i> 89, 977-982.
718 719 720 721	Nottin S, Vinet A, Stecken F, N'Guyen LD, Ounissi F, Lecoq AM & Obert P. (2002). Central and peripheral cardiovascular adaptations to exercise in endurance-trained children. <i>Acta Physiol Scand</i> 175 , 85-92.
722 723 724 725	Obert P, Mandigout S, Vinet A, N'Guyen LD, Stecken F & Courteix D. (2001). Effect of aerobic training and detraining on left ventricular dimensions and diastolic function in prepubertal boys and girls. <i>Int J Sports Med</i> 22 , 90-96.
726 727 728	Obert P, Mandigouts S, Nottin S, Vinet A, N'Guyen LD & Lecoq AM. (2003). Cardiovascular responses to endurance training in children: effect of gender. <i>Eur J Clin Invest</i> 33 , 199-208.
729 730 731 732	Obert P, Stecken F, Courteix D, Lecoq AM & Guenon P. (1998). Effect of long-term intensive endurance training on left ventricular structure and diastolic function in prepubertal children. <i>Int J Sports Med</i> 19, 149-154.
733 734 735 736	Pelliccia A, Maron BJ, Culasso F, Spataro A & Caselli G. (1996). Athlete's heart in women. Echocardiographic characterization of highly trained elite female athletes. <i>JAMA</i> 276 , 211-215.
737 738 739	Pluim BM, Zwinderman AH, van der Laarse A & van der Wall EE. (2000). The athlete's heart. A meta- analysis of cardiac structure and function. <i>Circulation</i> 101 , 336-344.

740 741	Prior DL & La Gerche A. (2012). The athlete's heart. <i>Heart</i> 98, 947-955.
742 743 744	Prommer N & Schmidt W. (2007). Loss of CO from the intravascular bed and its impact on the optimised CO-rebreathing method. <i>Eur J Appl Physiol</i> 100 , 383-391.
745 746 747 748	Prommer N, Wachsmuth N, Thieme I, Wachsmuth C, Mancera-Soto EM, Hohmann A & Schmidt WFJ. (2018). Influence of Endurance Training During Childhood on Total Hemoglobin Mass. <i>Front Physiol</i> 9 , 251.
749 750 751	Ratel S & Blazevich AJ. (2017). Are Prepubertal Children Metabolically Comparable to Well-Trained Adult Endurance Athletes? <i>Sports Med</i> 47, 1477-1485.
752 753 754 755	Ratel S, Tonson A, Le Fur Y, Cozzone P & Bendahan D. (2008). Comparative analysis of skeletal muscle oxidative capacity in children and adults: a 31P-MRS study. <i>Appl Physiol Nutr Metab</i> 33, 720-727.
756 757 758	Remes K. (1979). Effect of long-term physical training on total red cell volume. <i>Scand J Clin Lab Invest</i> 39, 311-319.
759 760 761	Rosner B, Prineas RJ, Loggie JM & Daniels SR. (1993). Blood pressure nomograms for children and adolescents, by height, sex, and age, in the United States. <i>J Pediatr</i> 123 , 871-886.
762 763 764 765	Runacres A, Mackintosh KA & McNarry MA. (2019). The effect of constant-intensity endurance training and high-intensity interval training on aerobic and anaerobic parameters in youth. <i>J Sports Sci</i> 37 , 2492-2498.
766 767 768	Sawilowsky SS. (2009). New effect size rules of thumb. <i>Journal of Modern Applied Statistical Methods</i> 8, 597–599.
769 770 771	Schmidt W & Prommer N. (2005). The optimised CO-rebreathing method: a new tool to determine total haemoglobin mass routinely. <i>Eur J Appl Physiol</i> 95, 486-495.
772 773 774 775	Skattebo O, Bjerring AW, Auensen M, Sarvari SI, Cumming KT, Capelli C & Hallen J. (2020). Blood volume expansion does not explain the increase in peak oxygen uptake induced by 10 weeks of endurance training. <i>Eur J Appl Physiol</i> 120 , 985-999.
776 777 778 779	Slaughter MH, Lohman TG, Boileau RA, Horswill CA, Stillman RJ, Van Loan MD & Bemben DA. (1988). Skinfold equations for estimation of body fatness in children and youth. <i>Hum Biol</i> 60, 709-723.
780 781 782	Van Praagh E & Dore E. (2002). Short-term muscle power during growth and maturation. <i>Sports Med</i> 32, 701-728.

784 785	Wanne OP & Haapoja E. (1988). Blood pressure during exercise in healthy children. Eur J Appl Physiol Occup Physiol 58, 62-67.
786 787 788 789	Weiner RB, DeLuca JR, Wang F, Lin J, Wasfy MM, Berkstresser B, Stohr E, Shave R, Lewis GD, Hutter AM, Jr., Picard MH & Baggish AL. (2015). Exercise-Induced Left Ventricular Remodeling Among Competitive Athletes: A Phasic Phenomenon. <i>Circ Cardiovasc Imaging</i> 8.
790 791 792	Wood CL, Lane LC & Cheetham T. (2019). Puberty: Normal physiology (brief overview). Best Pract Res Clin Endocrinol Metab 33 , 101265.
793	
794	

Table 1. Participant characteristics and cardiorespiratory fitness

		Во	pys	Training status main effect	Training status posthoc t-tests (ET vs. UN)	Maturity status main effect	Maturity statu t-tests (pre-	vs.post-	Interaction (Training status X Maturity status)	Gi	rls	Training status main effect	Training stat		Maturity status main effect	Maturity stat	- vs. post-	Interaction (Training status X Maturity status)
Anthropometric Characteristics		Pre-PHV	Post-PHV		Pre-PHV Post-PHV		ET	UN		Pre-PHV	Post-PHV		Pre-PHV	Post-PHV		ET	UN	
Age (years)	ΕT	11.7 ± 1.7	15.9 ± 1.1	P = 0.520	P = 0.046 P = 0.09	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.011	10.6 ± 1.3	14.1 ± 1.4	P = 0.154	P = 0.122	P = 0.585	P < 0.0001	P < 0.0001	P < 0.000	P = 0.566
	UN	10.6 ± 1.6	16.0 ± 1.2	P = 0.520	(d = 0.672) $(d = 0.593)$		(d = 2.530)	(d = 3.893)	P = 0.011	10.0 ± 1.2	13.8 ± 1.7	P = 0.154	(d = 0.510)	(d = 0.170)		(d = 2.571)	(d = 2.581)	
Maturity offset (years)	ET	-2.1 ± 1.2	1.5 ± 1.0	P = 0.686	P = 0.121 $P = 0.30$	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.067	-1.3 ± 1.0	1.9 ± 1.1	P = 0.089	P = 0.115	P = 0.391	P < 0.0001	P < 0.0001	P < 0.000	P = 0.648
	UN	-2.7 ± 1.1	1.9 ± 1.1	F = 0.000	(d = 0.517) $(d = 0.364)$		(d = 3.288)	(d = 4.067)	F = 0.007	-1.8 ± 0.9	1.6 ± 1.1		(d = 0.522)	(d = 0.269)	F < 0.0001	(d = 3.068)	(d = 3.342)	F = 0.048
Height (cm)	ET	148.6 ± 11.8	175.4 ± 8.6	P = 0.608	P = 0.457 $P = 0.95$	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.559	143.6 ± 9.6	164.7 ± 6.6	P = 0.016	P = 0.061	P = 0.144	P < 0.0001	P < 0.0001	P < 0.000	P = 0.413
	UN	145.9 ± 10.1	175.6 ± 10.6	7 = 0.000	(d = 0.245) $(d = 0.019)$		(d = 2.564)	(d = 2.872)		137.6 ± 9.6	161.7 ± 6.4		(d = 0.624)	(d = 0.461)	7 < 0.0001	(d = 2.566)	(d = 2.995)	
Body mass (kg)	ET	38.9 ± 8.9	61.7 ± 9.7	P = 0.788	P = 0.913 $P = 0.79$	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.901 34.4 ± 6.1	54.0 ± 8.5	P = 0.922	P = 0.649	P = 0.876	P < 0.0001	P < 0.0001	P < 0.000	P = 0.719	
	UN	39.3 ± 9.4	62.6 ± 10.3	7 - 0.700	(d = 0.036) $(d = 0.090)$		(d = 2.450)	(d = 2.373)		33.6 ± 5.5	54.5 ± 12.1	7 - 0.022	(d = 0.148)	(d = 0.049)		(d = 2.645)	(d = 2.186)) -0.713
Lean body mass (kg)	ET	33.2 ± 7.3	53.8 ± 7.1	P = 0.066	P = 0.114 $P = 0.30$	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.762	43.3 ± 6.0	P = 0.015	P = 0.081	P = 0.078	P < 0.0001	P < 0.0001	P < 0.000	P = 0.699	
	UN	29.8 ± 5.0	51.3 ± 6.2	7 - 0.000	(d = 0.526) $(d = 0.363)$		(d = 2.860)	(d = 3.814)		39.9 ± 6.2	7 = 0.010	(d = 0.580)	(d = 0.561)	7 < 0.0001	(d = 2.815)	(d = 2.713)		
Blood Pressure																		
Systolic BP (mm Hg)	ET	104 ± 8	117 ± 9	P = 0.502	P = 0.394 $P = 0.89$	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.628	104 ± 8	111 ± 7	P = 0.131	P = 0.218	P = 0.385	P < 0.0001	P = 0.003	P = 0.001	P = 0.725
	UN	102 ± 8	116 ± 7		(d = 0.284) $(d = 0.044)$)	(d = 1.454)	(d = 1.864)		101 ± 6	109 ± 6		(d = 0.416)	(d = 0.272)		(d = 0.958)	(d = 1.292)	.)
Diastolic BP (mm Hg)	ET	60 ± 7	64 ± 7	P = 0.521	P = 0.117 $P = 0.55$	P = 0.565	P = 0.114	P = 0.536	P = 0.130	62 ± 7	64 ± 7	P = 0.955	P = 0.831	P = 0.886	P = 0.242	P = 0.323	P = 0.490	P = 0.799
	UN	64 ± 6	62 ± 7	7 - 0.02	(d = 0.528) $(d = 0.207)$) - 0.000	(d = 0.507)	(d = 0.225)		63 ± 5	64 ± 6		(d = 0.071)	(d = 0.045)		(d = 0.302)	(d = 0.237)	
Cardiorespiratory Fitness																		
HR _{max} (beats·min)	ET	191 ± 9	194 ± 11	D 0000	P = 0.771 P = 0.27	D 0.050	P = 0.346	P = 0.080	D 0.500	196 ± 8	191 ± 6	D 0.450	P = 0.313	P = 0.302	D 0040	P = 0.031	P = 0.155	5
	UN	192 ± 9	197 ± 8	P = 0.303	(d = 0.096) $(d = 0.387)$	P = 0.056	(d = 0.296)	(d = 0.652)	P = 0.520 198 ± 7	198 ± 7	194 ± 10	P = 0.150	(d = 0.336)	(d = 0.324)	P = 0.013	(d = 0.667)	(d = 0.494)	P = 0.945
$\dot{V}O_{2max}\left(mL\cdot kg\cdot min\right)$	ET	59.4 ± 5.9	64.2 ± 8.0	P < 0.0001	P < 0.0001 P < 0.000	1 P = 0.017	P = 0.029	P = 0.217	P = 0.497	51.1 ± 6.3	51.7 ± 5.2	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.214	P = 0.737	P = 0.052	2
	UN	45.2 ± 7.6	48.0 ± 3.8	F < 0.0001	(d = 2.136) (d = 2.51		(d = 0.704)	(d = 0.454)	P = 0.497	41.9 ± 5.6	41.9 ± 5.6 38.1 ± 5.4	r < 0.0001	(d = 1.549)	(d = 2.573)		(d = 0.101)	(d = 0.685)	P = 0.093
$\dot{V}O_{2max}\left(\text{mL-kg}^{LBM}\text{-min}\right)$	ET	69.3 ± 6.1	73.2 ± 7.6	P < 0.0001	P < 0.0001 P < 0.000	1 P = 0.219	P = 0.070	P = 0.962	B = 0.104	62.0 ± 5.7	64.1 ± 5.2	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.954	P = 0.204	P = 0.275	P = 0.093
	UN	58.3 ± 6.4	58.2 ± 5.4	r < 0.0001	(d = 1.777) $(d = 2.235)$) = 0.219	(d = 0.577)	(d = 0.017)		P = 0.194 53.5 ± 5.3	51.2 ± 6.4		(d = 1.549)	(d = 2.235)		(d = 0.385)	(d = 0.377)	
$\dot{V}O_{2\text{max}}$ (% age predicted) ET	127 ± 13	145 ± 18	P < 0.0001	P < 0.0001 P < 0.000	P < 0.0001	P = 0.0004		P = 0.344	112 ± 14	139 ± 14	P < 0.0001		P < 0.0001	P < 0.0001	P < 0.0001		P = 0.010
	UN	97 ± 16	108 ± 9		(d = 2.136) $(d = 2.51)$) , , , , , , , , , , , , , , , , , , ,	(d = 1.198)	(d = 0.900)	7 - 0.0-17	92 ± 12	103 ± 14		(d = 1.549)	(d = 2.573)		(d = 1.963)	(d = 0.800)	7 = 0.010

<u>Key</u>: *BP*, blood pressure; *ES*, effect size; *ET*, endurance trained; $\dot{V}O_{2max}$, maximal heart rate; *PHV*, peak height velocity; *UT*, untrained; $\dot{V}O_{2max}$, maximal oxygen uptake.

Data expressed as mean \pm SD. Statistical comparisons were performed using two-way ANOVAs with training and maturity status as fixed factors. Independent samples *t*-tests were then used to identify differences where an interaction or main effect existed. Effect sizes calculated using Cohen's *d*. Participants for anthropometric characteristics included boys, pre-PHV (trained, n = 23 vs. untrained, n = 16) and post-PHV (trained, n = 19). Untrained, n = 19 vs. untrained vs. un

Group n's did not change from those of anthropometric characteristics, aside from the following: blood pressure, pre-PHV boys (trained, n = 22), pre-PHV girls (trained, n = 21 and untrained, n = 16); cardiorespiratory fitness, pre-PHV girls (untrained, n = 16).

Table 2. Left ventricular and haematological parameters in boys

					Training status posthers		•• • • • • •	Maturity state	us posthoc	Interaction	
		Во	ys	Training status main effect	Training status t-tests (ET v		Maturity status main effect	t-tests (pre	- vs. post-	(Training status X	
				main check	•		mum cricot	PH	V)	Maturity status)	
Absolute LV Parameters		Pre-PHV	Post-PHV			Post-PHV		ET	UN		
IVSd (mm)	ET	5.4 ± 1.3	7.4 ± 1.3	P = 0.028		P = 0.033	P < 0.0001	P < 0.0001		P = 0.328	
	UN	5.1 ± 0.7	6.6 ± 0.7		(d = 0.301) (d			(d = 1.488)			
LVIDd (mm)	ET	42.2 ± 4.3	50.2 ± 3.8	P = 0.029	P = 0.087 F		P < 0.0001	P < 0.0001		P = 0.930	
	UN	40.0 ± 2.9	47.9 ± 5.7		(d = 0.572) (d	t = 0.493		(d = 1.981)	(d = 1.752)		
LVPWd (mm)	ET	5.9 ± 1.5	7.8 ± 1.0	P = 0.036	P = 0.638 P		P < 0.0001	P < 0.0001		P = 0.165	
	UN	5.7 ± 0.9	6.9 ± 0.8		(d = 0.155) (d	,		(d = 1.485)			
LV length (cm)	ET	7.1 ± 0.8	8.5 ± 0.6	P < 0.0001	P = 0.0003 P		P < 0.0001	P < 0.0001		P = 0.807	
	UN	6.2 ± 0.5	7.6 ± 0.7		(d = 1.290) (d			(d = 2.005)			
LV mass (g)	ET	88.5 ± 25.0	155.6 ± 26.5	P < 0.0001	P = 0.027 P		P < 0.0001	P < 0.0001		P = 0.106	
	UN	73.2 ± 10.2	123.1 ± 20.1		(d = 0.749) (d			(d = 2.611)			
Relative wall thickness	ET	0.27 ± 0.06	0.30 ± 0.05	P = 0.509		P = 0.226	P = 0.044	P = 0.047		P = 0.287	
	UN	0.27 ± 0.04	0.28 ± 0.04		(d = 0.053) (d			(d = 0.634)			
EDV (mL)	ET	65.4 ± 17.5	104.2 ± 15.7	P < 0.0001	P = 0.002 P		P < 0.0001	P < 0.0001		P = 0.343	
	UN	49.5 ± 8.4	81.6 ± 15.0		(d = 1.092) (d			(d = 2.327)			
ESV (mL)	ET	26.7 ± 6.8	42.6 ± 6.6	P < 0.0001	P = 0.001 P		P < 0.0001	P < 0.0001		P = 0.281	
	UN	19.5 ± 4.3	32.1 ± 7.8		(d = 1.198) (d			(d = 2.365)			
SV (mL)	ET	38.7 ± 11.1	61.6 ± 10.8	P < 0.0001	P = 0.007 P		P < 0.0001	P < 0.0001		P = 0.451	
	UN	30.1 ± 5.4	49.5 ± 8.9		(d = 0.935) (d			(d = 2.089)			
Heart rate (beats·min)	ET	66 ± 12	51 ± 5	P = 0.001	P = 0.369 P		P < 0.0001	P < 0.0001		P = 0.037	
	UN	69 ± 9	64 ± 10		(d = 0.296) (d	,		(d = 1.520)			
Q (litres·min)	ET	2.36 ± 0.43	3.16 ± 0.68	P = 0.209		P = 0.909	P < 0.0001	P < 0.0001		P = 0.318	
	UN	2.08 ± 0.37	3.13 ± 0.59		(d = 0.697) (d	t = 0.040		(d = 1.435)	(d = 2.165)		
Relative LV Parameters											
IVSd/height (mm·m)	ET	3.6 ± 0.6	4.2 ± 0.8	P = 0.038	P = 0.501 P		P = 0.005	P = 0.011		P = 0.220	
	UN	3.5 ± 0.4	3.7 ± 0.4		(d = 0.221) (d			(d = 0.832)			
LVIDd/height (mm·m)	ET	28.4 ± 2.4	28.7 ± 2.2	P = 0.044	P = 0.221 F	P = 0.114	P = 0.976	P = 0.737	P = 0.817	P = 0.692	
	UN	27.5 ± 2.1	27.3 ± 2.8		(d = 0.405) (d	d = 0.561		(d = 0.105)	(d = 0.084)		
LVPWd/height (mm·m)	ET	3.9 ± 0.8	4.5 ± 0.6	P = 0.068	P = 0.907 P	P = 0.009	P = 0.079	<i>P</i> = 0.017	P = 0.939	P = 0.098	
	UN	3.9 ± 0.6	3.9 ± 0.6	7 = 0.000	(d = 0.038) (d	d = 0.960	0.070	(d = 0.769)	(d = 0.028)	. = 0.000	
LV length/height (cm·m)	ET	4.8 ± 0.3	4.9 ± 0.3	P < 0.0001	P < 0.0001 P	= 0.0001	P = 0.317	P = 0.304	P = 0.644	P = 0.833	
	UN	4.3 ± 0.3	4.3 ± 0.4	. 10.0001	(d = 1.631) (d	d = 1.495	0.011	(d = 0.323)	(d = 0.168)	7 = 0.000	
SV/LBM (mL·kg)	ET	1.16 ± 0.20	1.15 ± 0.12	P = 0.0002	P = 0.033 P	P = 0.001	P = 0.377	P = 0.740	P = 0.383	P = 0.648	
	UN	1.02 ± 0.19	0.97 ± 0.15	. = 0.0002	(d = 0.719) (d	d = 1.282	0.0	(d = 0.104)	(d = 0.318)	7 = 0.0.10	
Q/LBM (mL·kg ^{LBM} , min)	ET	72.20 ± 9.90	58.59 ± 8.05	P = 0.751	P = 0.693 F	P = 0.385	P < 0.0001	<i>P</i> < 0.0001	P = 0.075	P = 0.377	
	UN	70.69 ± 13.86	61.77 ± 12.93	7 = 0.701	(d = 0.129) (d	d = 0.304	7 4 0.0001	(d = 1.494)	(d = 0.665)	7 = 0.077	
Haematological parameters											
Hb mass (g)	ET	449 ± 112	770 ± 120	P = 0.007	P = 0.198 P	P = 0.017	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.304	
	UN	400 ± 86	663 ± 122	. = 0.00.	(d = 0.468) (d	d = 0.887		(d = 2.777)	(d = 2.461)	. – 0.00 .	
Hb mass/BM (g·kg)	ET	11.6 ± 1.5	12.5 ± 1.1	P < 0.0001	P = 0.070 P	< 0.0001	P = 0.109	P = 0.032	P = 0.729	P = 0.296	
	UN	10.4 ± 2.0	10.6 ± 0.6		(d = 0.666) (d	t = 2.033		(d = 0.690)	(d = 0.138)		
Blood volume (mL)	ET	3742 ± 920	6084 ± 860	P = 0.001	P = 0.169 P	P = 0.002	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.184	
	UN	3326 ± 613	5113 ± 764		(d = 0.501) (d	d = 1.183)		(d = 2.621)	(d = 2.555)		
Blood volume/BM (mL·kg)	ET	96.8 ± 13.6	99.3 ± 9.5	P < 0.0001	P = 0.070 P	< 0.0001	P = 0.749	P = 0.495	P = 0.354	P = 0.246	
	UN	87.2 ± 15.9	82.7 ± 7.2		(d = 0.667) (d	d = 1.930)		(d = 0.214)	(d = 0.372)		
Plasma volume (mL)	ET	2399 ± 593	3775 ± 536	P = 0.001	P = 0.203 P		P < 0.0001	P < 0.0001		P = 0.135	
	UN	2151 ± 397	3130 ± 477		(d = 0.462) (d			(d = 2.424)	(d = 2.215)		
Plasma volume/BM (mL·kg)	ET	62.1 ± 9.2	61.7 ± 6.6	P = 0.0001	P = 0.104 P	< 0.0001	P = 0.152	P = 0.880	P = 0.099	P = 0.211	
	UN	56.3 ± 10.3	50.8 ± 6.0	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	(d = 0.595) (d	d = 1.714)		(d = 0.047)	(d = 0.675)		
Hb (g·dL)	ET	13.4 ± 1.0	14.1 ± 0.7	P = 0.737	P = 0.594 F	P = 0.279	P < 0.0001	P = 0.007	P = 0.001	P = 0.260	
	UN	13.2 ± 0.8	14.4 ± 0.9	22.	(d = 0.192) (d	t = 0.388		(d = 0.877)	(d = 1.462)		
Hct (%)	ET	39.4 ± 2.1	41.7 ± 1.9	P = 0.800		P = 0.272	P < 0.0001	P = 0.001		P = 0.165	
	UN	38.8 ± 1.9	42.6 ± 2.7		(d = 0.304) (d	d = 0.394		(d = 1.117)	(d = 1.567)		

<u>Key:</u> *BM*, body mass; *EDV*, end-diastolic volume; *ES*, effect size; *ESV*, end-systolic volume; ET, endurance trained; *Hb*, haemoglobin; *Hct*, haematocrit; *IVSd*, interventricular septum diastole; *LV*, left ventricle; *LBM*, lean body mass; *LVIDd*, LV internal diameter diastole; *ET*, endurance trained; *LVPWd*, LV posterior wall diastole; *PHV*, peak height velocity; *SV*, stroke volume; *Q*, cardiac output; *UT*, untrained.

Data expressed as mean \pm SD. Statistical comparisons were performed using two-way ANOVAs with training and maturity status as fixed factors. Independent samples *t*-tests were then used to identify differences where an interaction or main effect existed. Effect sizes calculated using Cohen's *d*. Participants for cardiac parameters included boys, pre-PHV (trained, n = 23 vs. untrained, n = 16) and post-PHV (trained, n = 19 vs. untrained, n = 15). Paricipants for haematological parameters included boys, pre-PHV (trained, n = 23 vs. untrained, n = 12) and post-PHV (trained, n = 19 vs. untrained, n = 14).

Table 3. Left ventricular and haematological parameters in girls

		Gi	rls	Training status main effect		tus posthoc ET vs. UN)	Maturity status main effect	t-tests (pr	tus posthoc e- vs. post- IV)	Interaction (Training status X Maturity status)
Absolute LV Parameters		Pre-PHV	Post-PHV		Pre-PHV	Post-PHV		ET	UN	
IVSd (mm)	ET	5.3 ± 1.0	6.7 ± 1.2	P < 0.0001	P = 0.007	P = 0.001	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.415
	UN	4.5 ± 0.6	5.6 ± 0.8	P < 0.0001	(d = 0.929)	(d = 1.146)		(d = 1.297)	(d = 1.510)	P = 0.415
LVIDd (mm)	ET	40.6 ± 3.4	45.5 ± 3.3	P = 0.010	P = 0.106	P = 0.046	P < 0.0001	P < 0.0001	P = 0.001	P = 0.705
	UN	38.8 ± 3.2	43.1 ± 4.1	F = 0.010	(d = 0.535)	(d = 0.645)	F < 0.0001	(d = 1.443)	(d = 1.162)	F = 0.703
LVPWd (mm)	ET	5.7 ± 1.0	7.1 ± 1.3	P < 0.0001	P = 0.016	P < 0.0001	P < 0.0001	P = 0.0002	P = 0.014	P = 0.046
	UN	5.0 ± 0.7	5.5 ± 0.6		(d = 0.812)	(d = 1.533)	F < 0.0001	(d = 1.244)	(d = 0.860)	F = 0.040
LV length (cm)	ET	6.4 ± 0.7	7.5 ± 0.6	P = 0.001	P = 0.091	P = 0.005	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.346
	UN	6.0 ± 0.4	6.9 ± 0.6	7 = 0.001	(d = 0.561)	(d = 0.940)	7 < 0.0001	(d = 1.717)	(d = 1.642)	7 = 0.540
LV mass (g)	ET	80.2 ± 16.2	126.0 ± 30.5	P = 0.0002	P = 0.011	P = 0.003	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.193
	UN	67.7 ± 11.6	101.1 ± 18.0	7 = 0.0002	(d = 0.867)	(d = 0.975)	7 (0.0001	(d = 1.873)	(d = 2.176)	7 = 0.100
Relative w all thickness	ET	0.27 ± 0.05	0.30 ± 0.04	P = 0.001	P = 0.098	P = 0.002	P = 0.053	P = 0.034	P = 0.333	P = 0.833
	UN	0.25 ± 0.04	0.26 ± 0.04		(d = 0.548)	(d = 1.053)		(d = 0.660)	(d = 0.328)	
EDV (mL)	ET	48.9 ± 11.4	75.7 ± 13.3	P < 0.0001	P = 0.089	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.0003	P = 0.030
	UN	42.9 ± 9.5	57.8 ± 12.4		(d = 0.565)	(d = 1.384)		(d = 2.155)	(d = 1.341)	
ESV (mL)	ET	18.1 ± 4.7	30.0 ± 6.8	P < 0.0001	P = 0.308	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.014	P = 0.003
	UN	16.7 ± 3.9	21.0 ± 5.8		(d = 0.334)	(d = 1.421)		(d = 2.049)	(d = 0.862)	
SV (mL)	ET	30.8 ± 7.9	45.7 ± 7.8	P = 0.0001	P = 0.054	P = 0.001	P < 0.0001	P < 0.0001	P < 0.0001	P = 0.202
	UN	26.2 ± 5.8	36.8 ± 7.4		(d = 0.643)	(d = 1.159)		(d = 1.886)	(d = 1.585)	
Heart rate (beats·min)	ET	72 ± 10	61 ± 7	P = 0.001	P = 0.104	P = 0.0002	P < 0.0001	P < 0.0001	P = 0.034	P = 0.681
	UN	79 ± 16	70 ± 7		(d = 0.538)	(d = 1.275)			(d = 0.736)	
Q (litres·min)	ET	2.17 ± 0.60	2.79 ± 0.49	P = 0.117	P = 0.572	P = 0.089	P < 0.0001	P = 0.001	P = 0.024	P = 0.529
	UN	2.07 ± 0.56	2.50 ± 0.55		(d = 0.184)	(d = 0.546)		(d = 1.119)	(d = 0.787)	
Relative LV Parameters										
VSd/height (mm·m)	ET	3.7 ± 0.6	4.1 ± 0.6	P = 0.0001	P = 0.041	P = 0.001	P = 0.040	P = 0.040	P = 0.395	P = 0.333
	UN	3.3 ± 0.6	3.5 ± 0.4		, ,	(d = 1.124)			(d = 0.288)	
LVIDd/height (mm·m)	ΕT	28.3 ± 2.3	27.6 ± 1.6	P = 0.277		P = 0.137	P = 0.019		P = 0.041	P = 0.362
	UN	28.2 ± 2.0	26.7 ± 2.4		, ,	(d = 0.475)			(d = 0.708)	
LVPWd/height (mm·m)	ΕT	4.0 ± 0.6	4.3 ± 0.7	P < 0.0001		P < 0.0001	P = 0.587		P = 0.143	P = 0.033
	UN	3.7 ± 0.5	3.4 ± 0.3		,	(d = 1.552)			(d = 0.501)	
LV length/height (cm·m)	ET	4.4 ± 0.4	4.5 ± 0.3	P = 0.049		P = 0.011	P = 0.966		P = 0.269	P = 0.134
	UN	4.4 ± 0.3	4.3 ± 0.4		, ,	(d = 0.837)			(d = 0.375)	
SV/LBM (mL·kg)	ET	1.09 ± 0.21	1.06 ± 0.16	P = 0.010		P = 0.006	P = 0.138		P = 0.052	P = 0.520
Off Date () IRM ;)	UN	1.01 ± 0.12	0.92 ± 0.14		,	(d = 0.907)			(d = 0.672)	
Q/LBM (mL·kg ^{LBM} . min)	ET	76.66 ± 14.60		P = 0.815	P = 0.497	P = 0.553	P < 0.0001		P = 0.001	P = 0.364
Harmatala elada anamatana	UN	80.19 ± 17.57	62.84 ± 10.96		(a = 0.222)	(d = 0.188)		(a = 0.902)	(d = 1.201)	
Haematological parameters	_	254 . 70	F27 - 02		D 0.046	D 0.004		D . 0 0004	P < 0.0001	
Hb mass (g)	ET	351 ± 70	527 ± 82	P < 0.0001	P = 0.016	P = 0.001	P < 0.0001			P = 0.480
Lib mana/DM/milm)	UN	285 ± 73	434 ± 89			(d = 1.093)			(d = 1.799)	
Hb mass/BM (g·kg)	ET	10.3 ± 1.5	10.0 ± 1.4	P < 0.0001		P = 0.0001	P = 0.562		P = 0.894	P = 0.739
Blood volume (mL)	UN	8.2 ± 1.4	8.1 ± 1.2			(d = 1.370)			(d = 0.050)	
Blood volume (mL)	ET	2979 ± 575 2550 ± 776	4459 ± 580 3647 ± 788	P = 0.0003		P = 0.001 ($d = 1.193$)	P < 0.0001	P < 0.0001	P = 0.001 $(d = 1.401)$	P = 0.238
Blood volume/BM (mL·kg)	UN	87.6 ± 12.9				(u = 1.193) $P < 0.0001$			P = 0.289	
blood volume/blvi (the kg)	ET	73.2 ± 15.9	84.4 ± 10.9 68.1 ± 9.8	P < 0.0001		(d = 1.561)	P = 0.165		(d = 0.403)	P = 0.749
Plasma valuma (ml.)	UN	1941 ± 371	2855 ± 371			P = 0.006		P < 0.0001		
Plasma volume (mL)	ET	1941 ± 371 1693 ± 548	2835 ± 371 2416 ± 572	P = 0.003		P = 0.006 $(d = 0.931)$	P < 0.0001		P = 0.002 ($d = 1.284$)	P = 0.391
Plasma volume/BM (mL·kg)	UN	57.1 ± 8.5	54.1 ± 7.1			(a = 0.931) P = 0.003			P = 0.435	
i idoma voidine/Divi (IIIL'NY)	ET	57.1 ± 8.5 48.5 ± 11.2	54.1 ± 7.1 45.4 ± 10.2	P = 0.0002		P = 0.003 $(d = 1.013)$	P = 0.159		P = 0.435 $(d = 0.295)$	P = 0.987
Hb (g·dL)	UN		45.4 ± 10.2 13.1 ± 0.7		,	P = 0.650			(a = 0.295) $P = 0.021$	
I ID (g UL)	ET	12.9 ± 0.6 12.5 ± 0.9	13.1 ± 0.7 13.2 ± 0.7	P = 0.412		P = 0.650 $(d = 0.145)$	P = 0.008		P = 0.021 ($d = 0.888$)	P = 0.150
Het (%)	UN	38.3 ± 1.6	13.2 ± 0.7 39.5 ± 2.3			P = 0.698			P = 0.006	
Hct (%)	ET			P = 0.148			P = 0.001			P = 0.371
	UN	37.2 ± 1.8	39.2 ± 2.0		(a = 0.005)	(d = 0.124)		(a = 0.020)	(d = 1.070)	

<u>Key</u>: *BM*, body mass; *EDV*, end-diastolic volume; *ES*, effect size; *ESV*, end-systolic volume; ET, endurance trained; *Hb*, haemoglobin; *Hct*, haematocrit; *IVSd*, interventricular septum diastole; *LV*, left ventricle; *LBM*, lean body mass; *LVIDd*, LV internal diameter diastole; *ET*,

endurance trained; *LVPWd*, LV posterior wall diastole; *PHV*, peak height velocity; *SV*, stroke volume; *Q*, cardiac output; *UT*, untrained.

Data expressed as mean \pm SD. Statistical comparisons were performed using two-way ANOVAs with training and maturity status as fixed factors. Independent samples *t*-tests were then used to identify differences where an interaction or main effect existed. Effect sizes calculated using Cohen's *d*. Participants for cardiac parameters included girls, pre-PHV (trained, n = 22 vs. untrained, n = 17) and post-PHV (trained, n = 22 vs. untrained, n = 19). Participants for haematological parameters included girls, pre-PHV (trained, n = 21 vs. untrained, n = 12) and post-PHV (trained, n = 22 vs. untrained, n = 18).

Table 4. Bivariate associations with $\dot{V}O_{2max}$ (mL·kg^{LBM}·min) using pooled trained and untrained z-score values

		Во	ys		Girls						
	Pre	-PHV	Pos	t-PHV	Pre	-PHV	Post-PHV				
_	r	<i>P</i> -value	r	<i>P</i> -value	r	<i>P</i> -value	r	<i>P</i> -value			
IVSd/height	0.14	0.389	0.45	0.007	0.41	0.010	0.47	0.002			
LVPWd/height	0.04	0.814	0.47	0.005	0.31	0.056	0.59	<0.0001			
LVIDd/height	0.25	0.119	0.41	0.018	0.14	0.386	0.19	0.229			
LV mass/LBM	0.30	0.066	0.65	<0.0001	0.35	0.034	0.46	0.002			
ESV/LBM	0.49	0.001	0.64	<0.0001	0.27	0.106	0.43	0.005			
EDV/LBM	0.49	0.001	0.69	<0.0001	0.39	0.014	0.43	0.006			
SV/LBM	0.43	0.006	0.60	0.0001	0.41	0.011	0.35	0.023			
Hb mass/LBM	0.16	0.373	0.54	0.001	0.49	0.004	0.48	0.002			
Blood volume/LBM	0.15	0.386	0.49	0.004	0.39	0.026	0.45	0.003			

<u>Key</u>: *EDV*, end-diastolic volume; *ESV*, end-systolic volume; *Hb*, haemoglobin; *IVSd*, interventricular septum diastole; *LBM*, lean body mass; *LV*, left ventricle; *LVIDd*, LV internal diameter diastole; *LVPWd*, LV posterior wall diastole; *PHV*, peak height velocity; *SV*, stroke volume.

Bivariate correlation analysis was performed to identify independent associations with $\dot{V}O_{2max}$ using pooled trained and untrained *z*-score values. Analysis of cardiac structural variables included boys, pre-PHV (total, n=39 (trained, n=23; untrained, n=16)) and post-PHV (total, n=34 (trained, n=19), and post-PHV (total, n=39). Analysis of Hb mass and blood volume included boys, pre-PHV (total, n=35) (trained, n=23; untrained, n=12)) and post-PHV (total, n=34) (trained, n=19); untrained, n=14)), girls, pre-PHV (total, n=33) (trained, n=21; untrained, n=12)) and post-PHV (total, n=34) (trained, n=12)) and post-PHV (total, n=34)).

Table 5. Regression analyses with $\dot{V}O_{2max}$ (mL·kg^{LBM}·min) as the dependent variable for each pre- and post-PHV group using trained and untrained pooled *z*-score values

Group	Model	b	r partial	<i>P</i> -value	<i>R</i> ² Change	R^2_{adj}	<i>P</i> -value	SE	Constant Equation
Pre-PHV boys	EDV/LBM	0.494	0.494	0.001	0.244	0.224	0.001	0.893	y = 0.494x - 0.0001
Post-PHV boys	EDV/LBM	0.516	0.620	0.0002	0.481	0.608	<0.0001	0.639	y = 0.516x + 0.295x 0.282x + 0.015
	IVSd/height	0.295	0.437	0.014	0.098				
	Hb mass/LBM	0.282	0.395	0.028	0.066				
Pre-PHV girls	Hb mass/LBM	0.427	0.478	0.007	0.243	0.317	0.002	.0.799	y = 0.489x + 0.413x + 0.013
	IVSd/height	0.336	0.394	0.028	0.118				
Post-PHV girls	LVPWd/height	0.607	0.613	<0.0001	0.339	0.490	<0.0001	0.772	y = 0.607x + 0.416x - 0.043
	Hb mass/LBM	0.416	0.519	0.001	0.178				

<u>Key</u>: *EDV*, end-diastolic volume; *Hb*, haemoglobin; *IVSd*, interventricular septum diastole; *LBM*, lean body mass; *LV*, left ventricle; *LVPWd*, LV posterior wall diastole; *PHV*, peak height velocity.

Stepwise multiple linear regression analyses were used to identify regressions models which best account for the variance in $\dot{V}O_{2max}$ using pooled trained and untrained *z*-score values. Analysis of cardiac structural variables included boys, pre-PHV (total, n=39 (trained, n=23; untrained, n=16)) and post-PHV (total, n=34 (trained, n=19; untrained, n=15)), girls, pre-PHV (total, n=39 (trained, n=22; untrained, n=19)). Analysis of Hb mass and blood volume included boys, pre-PHV (total, n=39) (trained, n=19) and post-PHV (total, n=39) (trained, n=19) and post-PHV (total, n=39) (trained, n=19), girls, pre-PHV (total, n=39) (trained, n=19), untrained, n=19) and post-PHV (total, n=39) (trained, n=19), untrained, n=19), and post-PHV (total, n=39) (trained, n=19)).

Abstract figure legend. Schematic diagram depicting cardiac structural and haematological differences between trained and untrained boys and girls, pre-peak height velocity (PHV) and post-PHV alongside cardiac and haematological variables contributions to the variance in $\dot{V}O_{2max}$. Cardiac and haematological variables are greater in trained vs. untrained pre-pubertal children, and a greater number and magnitude of differences are observed at post-PHV. These variables provide significant predictive models for maximal oxygen consumption in children and are much stronger post-PHV, suggesting that other important determinants within the oxygen transport chain could account for the majority of variance in $\dot{V}O_{2max}$ before puberty.

Figure 1. Endurance-trained vs. untrained between-group differences in left ventricular (LV) mass, end-diastolic volume (EDV), blood volume and haemoglobin (Hb) mass for boys and girls, pre-peak height velocity (PHV) and post-PHV. Statistical comparisons were performed using two-way ANOVAs with training and maturity status as fixed factors. Independent samples t-tests were then used to identify differences where a main effect or interaction existed. Effect sizes calculated using Cohen's d. Participants for LV mass and EDV comparisons included boys, pre-PHV (trained, n = 23 vs. untrained, n = 16) and post-PHV (trained, n = 19 vs. untrained, n = 15), girls, pre-PHV (trained, n = 22 vs. untrained, n = 19). Participants for blood volume and Hb mass comparisons included boys, pre-PHV (trained, n = 23 vs. untrained, n = 12) and post-PHV (trained, n = 14), girls, pre-PHV (trained, n = 21 vs. untrained, n = 12) and post-PHV (trained, n = 22 vs. untrained, n = 18).

Figure 2. Linear regression analysis between end-diastolic volume (EDV) and blood volume for boys and girls, pre-peak height velocity (PHV) (total boys, n = 35 (trained, n = 23; untrained, n = 12) and total girls, n = 33 (trained, n = 23; untrained, n = 10)) and post-PHV (total boys, n = 33 (trained, n = 19; untrained, n = 14) and total girls, n = 39 (trained, n = 21; untrained, n = 18)). Statistical significance on the figures are from the linear regression analyses to indicate slope significance, with the r^2 also reported to indicate the relationship strength.





